

NPHL/Ambulatory Surgery Center Specimen Collection Workgroup

Friday, July 28, 2023

MEETING Minutes

Call to Order	Karen brought the meeting to order at 0736am. The following were in attendance: Stacy Hudson, Beth Nilson, Kim Zieno, Janel Myers, Anna McGrain, Laura Friend, Janet Estrada, Karen Stiles.
Review minutes & approve agenda	Minutes and agenda can be found on www.nphl.org , under menu “Chemical Terrorism,” then “Community Preparedness.” Unable to review minutes during meeting due to technical errors. Therefore, they will be posted to the site.
Alert Training & Drill	Awaiting the State of NE DHHS HAN System.
ASC Contact List	No changes to the contact database
Full Scale Exercise Fall 2022 Real Event CoVID19 AAR Report	Karen summarized main bottlenecks in FSX, set-up, labeling of specimens and NUIrt. Training of specimen collection with be introduced at this meeting.
ASC Specimen Collection Training	Specimen collection & labels experienced higher error rate in FSX last fall compared to previous exercises, primarily that instructions were not properly distributes, nor training provided prior. We reviewed the following: <ol style="list-style-type: none"> 1. CDC collection instructions on NPHL.org website and talked through entire steps. (see attached). 2. Disinfection of all specimens with disinfection wipes, both specimens and biohazard bags in which specimens are stored. 3. When collecting blood, do not anchor vein above needle to prevent needle stick. This was captured in a photo in World Herald.
Tabletop Exercise 2023 – Set up	Focus for TTX will be “Set up for Specimen Collection TTX.” The Tabletop will be held in 2 parts (see below).
Drills, Exercises at ASC Facilities	No other ASC drills or exercises were mentioned this past month.
Future Training Other Comments, Concerns, Questions	<u>Plan 2023 Calendar – suggestions:</u> Red Preparedness Tote Bag – review purpose & equipment Bomb threat Surgical Fire in your fire drill Lipid Rescue (http://lipidrescue.org/) August 18 (New Date) -DHHS Inspection preparedness -Guest Speaker BJ Knoell Sept 22 – Tabletop Part 1 -Inventory @Zoom Oct 6 - Tabletop Part 2 -Specimen Collection Setup @Methodist
Adjourn	Adjourned @ 0819 Next meeting will be Friday – August 18, 2023 by Zoom

CDC Specimen-Collection Protocol for a Chemical-Exposure Incident

See "Chemical Agents: Shipping Instructions for Specimens Collected from People who May Have Been Exposed to Chemical Agents" http://emergency.cdc.gov/databases/specimens_shipping_instructions.asp

Collect blood and urine samples for each person involved in the chemical-exposure incident.

Note: For children, collect only urine samples unless otherwise directed by CDC.

Blood-Sample Collection

For each person, collect blood in glass or plastic tubes in the following order: 1st: collect specimens in three (3) EDTA (purple-top) 4 mL or larger plastic or glass tubes; 2nd: collect another specimen in one (1) gray- or green-top tube. Collect the specimens by following the steps below:

- 1** Collect a minimum of 12 mL of blood in three (3) 4 mL or larger glass or plastic tubes. If using 3 mL tubes, use four tubes.



Do not use gel separators.

- 2** Mix contents of tubes by inverting them 5 or 6 times.



Tube #1 Tube #2 Tube #3

Label tubes in order of collection, #1, #2, #3

- 3** Place bar-coded labels on each tube, so that when the tubes are upright, the barcodes look like a ladder.



Tube #1 Tube #2 Tube #3

Store samples at 4°C to 10°C. Do not freeze.

- 4** After collecting samples in the purple-top tubes, collect one (1) sample in a gray- or green-top tube (gray-top tube shown). Allow the tube to fill to its stated capacity.



Do not use gel separators.

- 5** Mix contents of the tube by inverting it 5 or 6 times.



- 6** Place bar-coded labels on the tube, so that when the tube is upright, the barcodes look like a ladder.



Store samples at 4°C to 10°C. Do not freeze.

Urine-Sample Collection

For each person, collect 40 mL - 60 mL of urine in a screw-cap urine cup.



Label the urine cup with the appropriate bar-coded label as shown. Indicate on the cup how the sample was collected (if the method was other than "clean catch" (i.e., catheterization)).

Freeze samples *loosely* at -70°C.



Place bar-coded labels on all cups, so that when the cup is upright, the barcodes look like a ladder.

